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TCDD requires retinoic acid to induce cleft palate

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2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD) and retinoic acid (RA) exposure in wild type mice at embryonic day (E) 10.5 synergistically induce cleft palate, suggesting a possible interaction between TCDD and RA signalling pathways. Using quantitative RT-PCR, we first showed that neither TCDD nor RA treatments altered expression of genes transducing the RA (i.e., *Raldh3*, *Rarg*, *Crabp2*, *Cyp26a1*) or AHR (i.e., *Ahr*, *Arnt*, *Arnt2*, *Ahrr*) signals, respectively. We then analysed palate formation following exposure to TCDD (30 microgramme/kg) in *Raldh3*^{-/-} and *Rarg*^{-/-} fetuses lacking the RA synthesizing enzyme RALDH3 or the RA receptor RAR γ , respectively. All of them were resistant to cleft palate, indicating the requirement of an intact, active, RA signalling pathway allowing TCDD to induce this malformation. Analysis of reporter activity driven by a RA-responsive transgene in *Raldh3*^{-/-} palates indicated that RALDH3 is the only provider of RA, while in situ hybridization showed that *Raldh3* and *Rarg* expressions are restricted to nasal epithelium and mesenchyme, respectively. This finding entails that RA synthesized by RALDH3 in nasal epithelium diffuse towards mesenchyme to activate RAR γ , which may control, together with AHR, downstream genes involved in palate formation. Accordingly, we found that *Msx1* expression, which is restricted to palatal mesenchyme and whose impairment through gene ablation yields cleft palate, is repressed in RA- or TCDD-treated fetuses. We additionally found that TCDD is less potent to reduce *Msx1* expression in *Raldh3*^{-/-} fetuses. Altogether, our data show that TCDD- and RA-signalling pathways converge to control *Msx1* expression, whose impairment likely yields cleft palate.